

Levels of Fructose-2,6-Bisphosphate in Lymphocytes of Diabetic Patients

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Abstract:

Background: Patients with diabetes mellitus have infections more often than those without diabetes. Several factors predispose diabetic patients to infections, including an alteration in immune defense mechanism. Elevated levels of fructose-2,6- biphosphate in lymphocyte have been shown in diabetic patients. The activation of glycolysis by fructose -2,6- biphosphate in peripheral blood mononuclear cells causes the accumulation of glycolytic metabolites and inhibits the activation of immune cells.

Objective: To observe and compare the levels of fructose-2,6-bisphosphate in lymphocytes of diabetics and normal subjects

Material and Methods: 200 diabetic and 50 control subjects were selected for study .The subjects were evaluated for severity of diabetes and their fasting blood glucose, HbA_{1c}, total leucocyte count, lymphocyte count and fructose-2,6-bisphosphate in lymphocytes were estimated.

Results: The results show that mean fasting blood glucose, HbA_{1c}, total leucocytes count and fructose-2,6-bisphosphate levels in lymphocytes were significantly higher ($P < 0.001$) while lymphocyte count was significantly lower ($P < 0.001$) in diabetic patients as compared to control group.

Conclusions: It was concluded from the facts observed in this study that elevated levels of fructose-2,6-bisphosphate in lymphocytes and decreased number of lymphocytes may have induced chances of infections in diabetic patients.

Keywords: Lymphocytes, Fructose-2,6-bisphosphate, Diabetes, Infections.

Introduction:

Fructose-2,6-bisphosphate is detected in all mammalian tissue¹. It is powerful allosteric activator of 6 phosphofructose-1-kinase which is the rate limiting enzyme for glycolysis². When levels of fructose-2,6-bisphosphate are high, glycolysis is enhanced and gluconeogenesis is inhibited³. Diabetes causes substantial changes in the fructose-2,6-bisphosphate system. In hepatocyte, diabetes mellitus enhances phosphorylation of fructose-2,6-bisphosphate leading to a decrease in the activity of the enzyme causing hyperglycemia. In peripheral blood lymphocytes fructose-2,6-bisphosphate system is slightly different from that of hepatocyte⁴. The activation of glycolysis by fructose-2,6-bisphosphate in peripheral blood mononuclear cells causes the accumulation of glycolytic metabolites and inhibits the activation of immune cells⁵. These altered metabolic products and oxidative stress play a role in the development of dia-

betic complications⁶. In vitro evidence shows that neutrophil function and humoral immunity may be depressed in people with diabetes⁷.

Hyperglycemia increases intracellular fructose-2,6-bisphosphate in immune cells⁸. Elevated level of fructose-2,6-bisphosphate in lymphocyte have been shown in diabetic patients. These findings suggest the association between accelerated glycolysis due to hyperglycemia and alteration of the immune system during the diabetic state⁹ and may help to determine the impaired function of immune cells in patients with diabetes¹⁰. Patients with diabetes mellitus have infections more often than those without diabetes. The course of infection is also more complicated in this patient group¹¹. Good metabolic control is a major factor in limiting the development and spread of infection¹²

Methodology

This study was carried out in the Department of Biochemistry, Basic Medical Sciences Institute (BMSI), Jinnah Postgraduate Medical Centre (JPMC) Karachi. The study protocol was approved by the Local Bioethical Committee and informed consent was obtained from all subjects. Total 250 individuals were included in the study, these were divided into two groups. Group A consist of 50 healthy individuals selected as control and group B consist of 200 diagnosed patients of diabetes mellitus of different age and sex with positive history of associated infection. Subjects suffering from anemia, liver diseases, renal diseases, any other endocrine dis-

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eases and pregnant women were excluded. Morning samples were taken after an overnight fast of at least 12-14 hours. About 10 ml blood was drawn from ante-cubital vein after all aseptic measures. 1 ml of blood was used to estimate HbA_{1c} by fast ion exchange resin separation method, using kit supplied by Human Germany Cat No 10658. 6ml of blood was heparinized for the separation of lymphocytes. Serum was separated from rest of the sample and used to estimate blood glucose by enzymatic calorimetric (GOD-PAP) method using kit, No.Cod.1001191 supplied by Spinreact, SA, Spain. Complete blood count (CBC) was done on Sysmex KX 21 automated cell counter, which measures and calculates total leucocyte and lymphocytes counts. Separation of lymphocytes from whole blood was done with lymphocyte separation medium (LSM) catlog no 25-072, Cl, 1×100 ml density 1.077-1.08 g/ml which is a separation solution made with Ficoll TM is a density gradient media and Dulbecco's phosphate buffered saline without Calcium and Magnesium Cat No 21040-CV. Ficoll TM is a hydrophilic polymer with a molecular weight of 400 Dalton. It is used for the production of density gradients for separation of cells and sub-cellular components, which sediment on centrifugation. Heparinized blood was centrifuged with LSM. Sedimented erythrocytes, polynuclear leukocytes and mononuclear lymphocytes were separated. Superficial lymphocyte layer was aspirated and washed with buffered balanced salt solution and resuspended in the appropriate medium for application.

Determination of fructose-2,6-bisphosphate in lymphocytes was done by chemical method based on the ability of fructose-2,6-bisphosphate to activate pyrophosphate dependent phosphofruktokinase from potato tubers Sigma Chemicals. Statistical analysis was performed using SPSS statistical software by paired student t-test.

Results

A total of two hundred and fifty subjects were studied. Group A consist of 50 healthy individuals as control and group B consist of 200 diabetic patients associated with infections.

Table-1 shows the comparison of the mean values of age, body mass index, total leukocyte count (TLC) and lymphocyte count between control and diabetic patients. It shows that mean body mass index, and TCL of diabetic patients were significantly higher ($P < 0.001$) as compared to control group. The mean value of lymphocyte count was significantly lower ($P < 0.001$) in diabetic group.

Table-2 Shows the comparison of the mean values of fasting blood glucose, HbA_{1c} and fructose-2,6-bisphosphate levels in lymphocytes between control and diabetic patients. The mean values of these variables were significantly higher ($P < 0.001$) in diabetic patients as compared to control.

Table-1

Comparison of Age, Body Mass Index, TLC and Lymphocyte Count of Control with the Diabetic Subjects.

The values are expressed as Mean \pm S.E.M. The number of observation and units are given in parenthesis. n = number of subjects.

Variables	Control n=50	Diabetic n=200
Age (Years)	44.90 \pm 1.36	43.26 \pm 0.56
BMI (kg/m ²)	24.12 \pm 0.45	27.63* \pm 0.31
TLC (10 ⁹ /L)	7.45 \pm 1.13	12.00* \pm 0.12
Lymphocyte count (%)	32.33 \pm 1.15	27.66* \pm 0.32

*.P < 0.001 significant when compared to control.

Table-2

Comparison of Fasting Blood glucose, HbA_{1c} and Fructose-2,6-bisphosphate in control and diabetics.

The values are expressed as Mean \pm S.E.M. the number of observation and units are given in parenthesis.

Variables	Control n=50	Diabetic n=200
FBS (mg/dl)	86.54 \pm 1.95	137.70* \pm 1.61
HbA _{1c} (%)	4.33 \pm 0.10	8.45* \pm 0.11
Fructose 2,6 bisphosphate (pmol)	3.15 \pm 0.11	6.91* \pm 0.11

n = number of subjects.

*.P < 0.001 significant when compared to control.

Discussion:

Diabetes is a group of metabolic diseases characterized by hyperglycemia that occurs when the pancreas does not produce enough insulin or body cannot effectively use the insulin it produces or both¹³. Infections tend to occur with greater frequency and severity in diabetic patients than in non diabetic. Several factors predispose diabetic patients to infections. These factors include: genetic susceptibility to infection, altered cellular and humoral immune defense mechanism, local factors including poor blood supply and nerve damage and alteration in metabolism associated with diabetes mellitus¹⁴. Specific defects in innate and adaptive immune function have been identified in many in vitro studies¹⁵. Our study shows that the mean fasting blood glucose, HbA_{1c} and fructose-2,6-bisphosphate levels in lymphocytes were significantly

higher in diabetic group. The results were in agreement with the other studies conducted on similar parameters^{4,5,10,16}. This data suggests that hyperglycemia increases fructose-2,6-bisphosphate in lymphocytes. Also in diabetic patients, significantly increased total leukocyte count and decreased lymphocyte count were observed in present study and this is in agreement with study carried out by earlier workers¹⁷.

It was concluded from the facts observed in this study that elevated levels of fructose-2,6-bisphosphate in lymphocytes and decreased number of lymphocytes may be responsible for the impaired function of immune cells and may have induced increased chances of infections in diabetic patients. Our analysis was based on a single measurement that may not reflect the relation over time. However there is a need for more well designed, randomized studies assessing the value of glycemic control and fructose-2,6-bisphosphate levels for better understanding of frequency of infections in diabetics.

Reference:

1. Louis HUE, Rider MH. Role of fructose 2,6-bisphosphate in the control of glycolysis in mammalian tissues. *Biochem.J* 1987; 245:313-24.
2. Okar DA, Manzano A, Navarro-Sabate A, Riera L, Bartrons R, Lange AJ. PFK-2/FBPase-2: maker and breaker of the essential biofactor fructose-2,6-bisphosphate. *Trends Biochem Sci* 2001;26: 30-35
3. Wu C, Khan SA, Peng LJ, Lange AJ. Roles for fructose-2,6-bisphosphate in the control of fuel metabolism: beyond its allosteric effects on glycolytic and gluconeogenic enzymes. *J Adv Enzyme Regual* 2006; 46: 72-88.
4. Belyaeva NF, Golubev MA, Markova MS, Col Chenko OL, Lamzina NN, Gorodetskii VK, Victorova LN, Balabolkin MI, and Korovkin BF. Determination of Fructose-2,6-bisphosphate in human lymphocyte is a potential laboratory test in *Diabetology* 1996; 122 (9) : 341-44.
5. Atsumi T, Chiba H, Yoshioka N, Bucala R, koike T. Increased fructose 2,6-bisphosphate in peripheral blood mononuclear cells of patients with diabetes. *Endocrine J* 2007;54(4): 517-20.
6. Brownlee M. The pathobiology of diabetic complications: a unifying mechanism. *Diabetes* 2005; 54: 1615-25.
7. Baiju R, Shah MD, Janet E, Hux MD. Quantifying the risk of infectious diseases for people with diabetes. *Diabetes Care* 2003; 26:510-13.
8. Vander Heiden M.G, Cantley L.C, Thampson C.B. Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science* 2009; 123:309-14.
9. Nabi AH, Islam LN, Rahman MM, Biswas KB. Polymorphonuclear neutrophil dysfunctions in streptozotocin- Induced type 1 diabetic rats. *JBiochemMolBiol* 2005;38:661-67.
10. Victoria R, Moreno - Auriolles, Montano R, Conde M, Bustos R , Sobrino F. Streptozotocin - Induced diabetes increases fructose 2,6-bisphosphate levels and glucose metabolism in thymus lymphocytes. *J Life Sciences* 1996;58: 477-84.
11. Geerlings SE, Hoepelman AI. Immune dysfunction in patients with diabetes mellitus. *FEMS Immunole Med Microbiol* 1999; 26(3-4):259-65.
12. Shanawany T, Williams PE, Jolles S. clinical immunology review series an approach to the patients with anaphylaxis. *Clin Exp Immunol* 2008;153(1):1-9
13. Expert Committee on Diagnostics and Classification of Diabetes Mellitus. Report of Expert Committee on Diagnostics and Classification of Diabetes Mellitus. *Diabetes Care* 2003; 26: S5 - 20.
14. Kornum JB, Thomson RW, Riis A, Levang H, Shonheyder H, Sorensen HI. Diabetes, glyce-mic control and risk of hospitalization with pneumonia. *Diabetes care* 2008;31(8):1541-45.
15. Peleg AY, James TW, Carthy, Timothy M, Davis E. Common infections in diabetes; Pathogenesis, management and relationship to gly-cemic control. *Diabetes/Metabolism Research and Reviews* 2007; 23(1): 3-13.
16. Bosca L, Mojena M, Jose M, Guerra D, Marquez C. Phorbol 12,13-dibutyrate and mitogens increase fructose 2,6 bisphosphate in lymphocyte. *Eur J Biochem* 1988;175:317-23.
17. Fu-Mei Chung,TsaiJC,Chang DM, Shin SJ, Lee YJ, Peripheral total and differential leukocyte count in diabetic nephropathy. *Diabetic care* 2005;28:1710-17.